A KEY BASED ON EPIDERMAL CHARACTERISTICS FOR THE IDENTIFICATION OF CERTAIN HIGHVELD GRASSES*

by

P. D. F. KOK** and H. P. VAN DER SCHIJFF**

Abstract – The grass leaf epidermis shows characteristic features which are taxonomically valuable and which renders leaf fragments identifiable in the stomach contents and faeces of animals. A brief account of the features of the grass epidermis is given, as well as the terminology which is used.

A key to identify 60 grass species of the Van Riebeeck Nature Reserve, near Pretoria, Republic of South Africa, is given. This key is based on abaxial leaf epidermal characteristics.

Introduction

The taxonomic significance and importance of the epidermis of the leaves of grasses have been dealt with by a large number of authors (Duval-Jouve, 1875; Pratt, 1932; Metcalfe, 1960). The characteristic taxonomic significance of the epidermis of grasses has been used to identify and classify different grass species. It has even been used to distinguish sub-species from one another (Goossens and Theron, 1934; Borrill, 1957). The most recent application of the epidermal identification of grass species is found in the identification of grass species in the stomach contents and faeces of animals (Baumgartner and Martin, 1939; Dusi, 1949; Martin, 1955, 1964; Davies, 1959; Hercus, 1960; Storr, 1961; Stewart, 1965, 1967, 1971; and Stewart and Stewart, 1970). In South Africa it is only during the past four years that an attempt has been made to identify food plants in the faeces of game (McAllister, 1967; Liversidge, 1970).

In the present study the epidermis of sixty grasses in the Van Riebeeck Nature Reserve, near Pretoria, Republic of South Africa, was studied in order to determine whether these species could be identified in the faeces of blesbok.

** Department of General Botany, University of Pretoria, Pretoria.

^{*}Part of the International Biological Research Programme (1967–1971). The research was supported by a grant from the I.B.P. through the South African Council for Scientific and Industrial Research and was presented by the senior author for the M.Sc. Degree at the University of Pretoria, Pretoria.

Material and Methods

Leaves from the top of the culm as well as from the tuft, of the different grass species were collected. Slides were prepared from the epidermis taken from the base, the middle, as well as from the apex of both kinds of leaves. The reason for preparing more than one slide for every grass species was to overcome intraspecific variations.

Various methods were used in the preparation of the slides (Croker, 1959; Hercus, 1960; Metcalfe, 1960; and Storr, 1961). Drawings of the epidermis were made with the aid of a Leitz S. M. microscope and Leitz

microprojector.

Features of the grass epidermis

A detailed description of the epidermis is given by Metcalfe (1960) and the terminology which he adopted was used to describe the epidermis of the different grasses. The following notes are intended to make the key intelligible and to give a general idea of the taxonomic features of the epidermis.

For descriptive purposes the epidermis is viewed with the long axis of the leaf horizontal in the field of the microscope. All the directions parallel to the long axis are then horizontal and those perpendicular to

the long axis vertical.

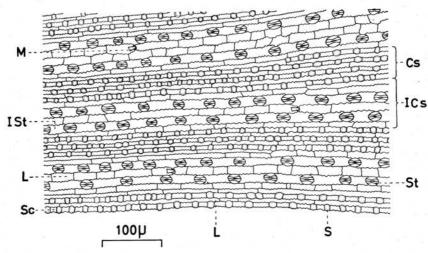


Fig. 1. Surface-view of the abaxial epidermis of *Tragus berteronianus* Schult. Cs costal zone; ICs intercostal zone; ISt interstomatal-cell; L long-cell; M micro-hair; S silica-cell; Sc short-cell; St stomata.

The epidermis of the grass leaf in surface view is divided horizontally into costal zones, which are characterized by the abundance of silicacells, and the intercostal zones between the costal zones (Fig. 1). According to Metcalfe (1960) two types of epidermis cells may be distinguished,

namely long-cells and short-cells. In this study three categories of cells, viz. long-cells, silica-cells and short-cells were identified.

1.0 Long-cells

These are cells with horizontal walls longer than twice the vertical walls. For identification purposes the shape of the cells and their wall were taken into consideration:

- (a) Long-cells with sinuous (Fig. 2A) and non-sinuous walls (Fig. 2B).
- (b) Brick-shaped long-cells (Fig. 2A) and inflated long-cells (Fig. 2B).

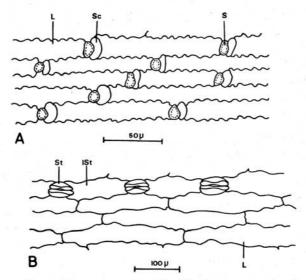


Fig. 2. Long-cells and interstomatal-cells. ISt interstomatal-cell; L long-cell; S silica-cell; Sc short-cell; St stomata.

2.0 Silica-cells

These cells contain a silica-body. The shape of the cell and the body do not always correspond. In the description of silica-cells it is important to state whether it is the shape of the cell or that of the body which is described (Metcalfe, 1960). In this study the description refers to the shape of the silica-body.

The following types of silica-bodies were recognized:

- (a) Dumb-bell shaped. Either long dumb-bell shaped (Fig. 3A) or short dumb-bell shaped (Fig. 3B).
- (b) Cross-shaped (Fig. 3C).
- (c) Nodular (Fig. 3D).
- (d) Saddle-shaped. The long axis can be either horizontal (Fig. 3E) or vertical (Fig. 3F).
- (e) Elliptical with the long axis horizontal (Fig. 3G) or vertical (Fig. 3H).
- (f) Crescent-shaped (Fig. 31).

NPB C

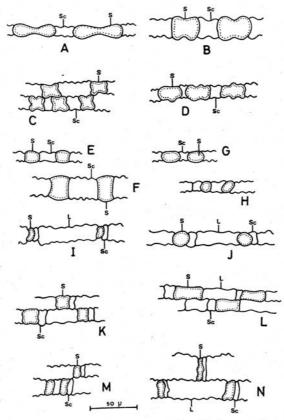
(g) Round (Fig. 3J).

(h) Quadrangular (Fig. 3K).

(i) Rectangular (Fig. 3L).

Tall and narrow (Fig. 3M).

(k) Irregular (Fig. 3N).



Silica-bodies. L long-cell; S silica-cell; Sc short-cell. Fig. 3.

3.0 Short-cells

The shapes illustrated for silica-cells are also common for the short-cells. In the costal and intercostal zones, the silica-cells are mostly accompanied by short-cells (Fig. 4).

4.0 Appendages of the epidermis

4.1 Micro-hairs

All two-celled hairs were regarded as micro-hairs to correspond with Metcalfe (1960). Based on the shape of the basal cell and the length of the distal cell, four types were dinstinguished:

(a) Basal cell tapering to the base (Fig. 5A).

(b) Basal cell not tapering towards the base (Fig. 5B).

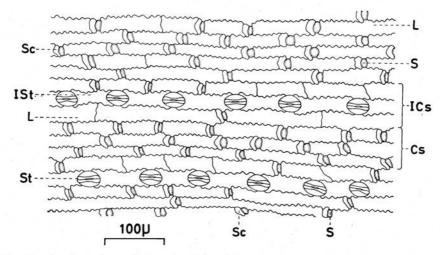


Fig. 4. Surface-view of the abaxial epidermis of *Eragrostis capensis* (Thunb.) Trin. Cs costal zone; ICs intercostal zone; ISt interstomatal-cell; L long-cell; S silica-cell; Sc short-cell; St stomata.

(c) Distal cell longer than the basal cell (Fig. 5C).

(d) Distal cell shorter than the basal cell (Fig. 5A).

Figs 5D and 5E show examples of one-celled micro-hairs which are too small to be regarded as macro-hairs.

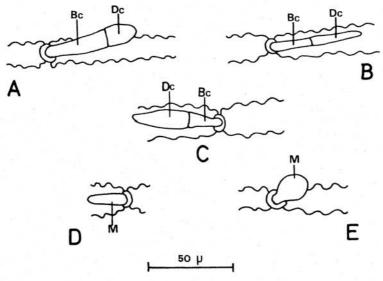


Fig. 5. Micro-hairs. Bc apical cell; Dc distal cell; M one-celled micro-hair.

4.2 Macro-hairs and prickle-hairs

According to Metcalfe (1960) it is sometimes difficult to distinguish between macro-hairs and prickle-hairs. The criterion used in this study is the length of the barb. The appendages with long barbs are regarded as macro-hairs and those with short barbs, as prickle hairs.

Macro-hairs

Two types of macro-hairs, based on the number of smaller epidermal cells surrounding the base, are distinguished:

(a) Base surrounded by one or two smaller epidermal cells (Fig. 6A).

(b) Base surrounded by more than two smaller epidermal cells (Fig. 6B).

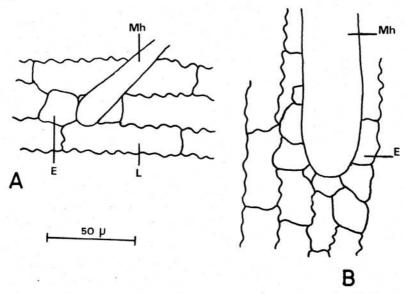


Fig. 6. Macro-hairs. E specialized (smaller) epidermal cell; L long-cell; Mh macro-hair.

Prickle-hairs

Based on the width of the base of the prickle-hairs, two types are recognized:

(a) Base narrower than adjacent cells (Fig. 7A). (b) Base broader than adjacent cells (Fig. 7C).

The shape of the base also differs and three types are distinguished:

(a) Round base (Fig. 7A).

(b) Quadrangular to rectangular base (Fig. 7B).

(c) Elliptical base (Fig. 7C).

4.3 Papillae

These can be divided into two groups according to the number of papillae per cell.

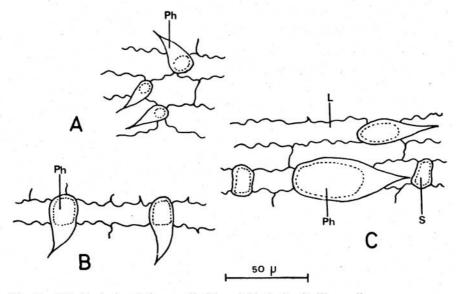


Fig. 7. Prickle-hairs. L long-cell; Ph prickle-hair; S silica-cell.

(a) One papilla per cell (Figs 8A and 8B).

(b) More than one papilla per cell (Fig. 8C). The sizes of the papillae vary (Figs 8A and 8B).

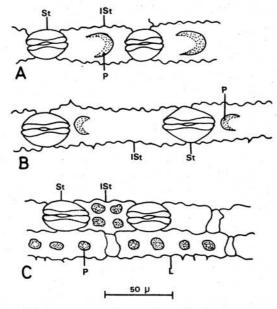
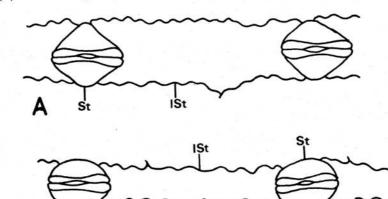


Fig. 8. Papillae. ISt interstomatal-cell; P papil; St stomata; L long-cell.

4.4 Subsidiary cells of the stomata

The shapes of the subsidiary cells vary and two types were recognized. However, this proved to be of minor taxonomic value.

(a) Triangular (Fig. 9A).(b) Dome-shaped (Fig. 9B).



50 H

Fig. 9. Subsidiary cells of stomata. ISt interstomatal cell; St stomata.

KEY

					•
1	Silica-bodies on costal zones dumb-bell shaped .			•	2
	Silica-bodies in costal zones not dumb-bell shaped				96
0	No long dumb-bell shaped silica-bodies in costal zone	S			3
4.	Long dumb-bell shaped silica-bodies in costal zones				49
	Cross shaped silica-hodies in costal zones	8	1	190	4
3.	(1055-Shaped Sinca Boards III		•	•	30
	No cross-shaped silica-bodies in costal zones		•	•	5
4.	Wolling - Shabed Sinea-bodies in costar	•	•	٠	
	No nodular-shaped silica-bodies in costal zones	•	•	•	16
5	Cells with papillae occur	•	٠		6
٥.	No cells with papillae occur				7
G	n iii land a sella ag avall ag an some inters	tom	ata	al-	
6.	cells	noch	loa	stag	nina
	cells	trun	2 50	บกรา	nsum
	Papillae on some interstomatal cells only Urely	itan	109	liano	nalic
7.	No short-cells and silica-cells in intercostal zones Dig	uar	iu u	iugo	0
	Short-cells and/or silica-cells in intercostal zones .	•	٠	•	0
8.	m · · · · · · · · · · · · · · · · · · ·	•	•	٠	9
•	Prickle-hairs absent	•	•		11
0	Prickle-hairs in costal and intercostal zones				10
9.	Prickle-hairs in costal zones only Rhynches	lytri	ım s	setifo	lium
	Prickie-mails in costal Zones only			,	

10.	Macro-hairs in intercostal zones only Urochloa panicoides
	Macro-hairs on the margin of the leaf only . Rhynchelytrum repens
11.	Macro-hairs occur
	Macro-hairs absent
12.	Micro-hairs occur
14.	Micro-hairs absent
13.	Majority of long-cells in intercostal zones alternate with a short-
15.	cell, silica-cell or a short- and silica-cell pair . Panicum laevifolium
	Minority of long-cells in intercostal zones alternate with a short-
	cell, silica-cell or a short- and silica-cell pair . <i>Urochloa panicoides</i>
1.4	Micro-hairs occur
14.	Micro-hairs occur
4.5	Micro-nairs absent
15.	Interstomatal-cells shorter than the length of the stomata Cymbopogon excavatus
	Interstomatal-cells longer than the length of the stomata Panicum laevifolium
16.	Tiregular shaped shired bodies occur an essential
00120	110 Hickural shaped since bodies occur
17.	Rows of long-cells between rows of silica-cells in costal zones
	Alloteropsis semialata
	No rows of long-cells between rows of silica-cells in costal zones
	Prickle-hairs occur
18.	Prickle-hairs occur
	Prickle-hairs absent
19.	Cells with papillae
	No cells with papillae
20.	Papillae on some long-cells, short-cells and interstomatal-cells
1	Echinochloa stagnina
	Papillae on some interstomatal-cells only . Diheteropogon amplectens
21.	Short-cells and silica-cells occur in intercostal zones 23
	No short-cells or silica-cells occur in the intercostal zones . 22
22.	Interstomatal-cells as long as or shorter than twice the length
	of the stomata Diheteropogon amplectens
	Interstomatal-cells longer than twice the length of the stomata
	Digitaria ternata
23.	Individual short-cells and silica-cells as well as short- and silica-
	cell pairs in the intercostal zones
	Only short- and silica-cell pairs in the intercostal zones 24
24.	
	long-cells between silica-cell rows in costal zone Diheteropogon
	amplectens
	Interstomatal-cells do not taper towards the stomata; long-
	cells occur between silica-cell rows in costal zones Setaria
	nigrirostris
25.	Prickle-hairs in costal zones only ,
7	Prickle-hairs in costal and intercostal zones 28

2	6.	Cells with papillae occur
_	_	No cells with papillae occur
2	7.	Short-cells and/or silica-cells occur in intercostal zones . Setaria nigrirostris
		No short-cells or silica-cells in intercostal zones . Digitaria ternata
0		Prickle-hairs mainly in the costal zones . Diheteropogon amplectens
2	8.	Prickle-hairs mainly on the sides of costal zones
0	0	Subsidiary-cells of stomata triangular Digitaria smutsii
2	9.	Subsidiary-cells of stomata dome-shaped Digitaria eriantha
2	20	Nodular-shaped silica-bodies in costal zones
J	30.	No nodular-shaped silica-bodies in costal zones
2	31.	Short- and/or silica-cells in intercostal zones
3	11.	No short- or silica-cells in intercostal zones
2	32.	Brick-shaped long-cells with sinuous walls Digitaria ternata
-	14.	Inflated long-cells with non-sinuous walls Setaria sphacelata
2	33.	Long-cells between silica-cell rows in costal zones 34
	,,,	No long-cells between silica-cell rows in costal zones Imperata
		cylindrica
9	34.	Individual short-cells and silica-cells as well as short- and silica-
٠	, 1.	cell pairs in intercostal zones Schizachyrium sanguineum
		Only short- and silica-cell pairs in intercostal zones 35
9	35.	Majority of long-cells in intercostal zones alternating with a
•		short- and silica-cell pair Setaria nigrirostris
		Minority (often none) of long-cells in intercostal zones alternate
		with a short- and silica-cell pair Setaria flabellata
5	36.	Prickle-hairs occur
		Prickle-hairs absent 43
:	37.	Prickle-hairs in intercostal zones only
		Prickle-hairs in costal and intercostal zones 39
	38.	Short-cells in intercostal zones Schizachyrium sanguineum
		No short-cells in intercostal zones Paspalum dilatatum
	39.	
		Macro-hairs absent
	40.	Macro-hairs in intercostal zones only Urochloa panicoides
		Macro-hairs on the margin of the leaf only . Rhynchelytrum repens
	41.	Short-cells and/or silica-cells in intercostal zones
		No short-cells or silica-cells in intercostal zones Digitaria diagonalis
	42.	Basal-cell of micro-hairs longer than stomata . Rhynchelytrum repens Basal-cell of micro-hairs shorter than stomata . Setaria woodii
		Dasar-cen of finero mans services
	43.	Macro-harrs occur
	44.	Majority of long-cells in intercostal zones alternate with a short- cell, silica-cell or a short- and silica-cell pair . Panicum laevifolium
		cell, silica-cell or a short- and silica-cell pair . Panicum laevifolium Minority of long-cells in intercostal zones alternate with a short-
		cell, silica-cell or with a short- and silica-cell pair <i>Urochloa panicoides</i>
	4=	
	45.	Long-cens between sinea-cen rows in costar zones

	No long-cells between silica-cell rows in costal zones Imperata
46.	Short-cells and/or silica-cells in the intercostal zones 47
	No short-cells or silica-cells in intercostal zones Digitaria diagonalis
47.	Interstomatal-cells longer than twice the length of the stomata,
	without papillae
	Interstomatal-cells shorter than twice the length of the stomata,
	and generally with papillae Schizachyrium sanguineum
48.	Majority of long-cells in intercostal zones alternate with a
	silica- and short-cell pair; one row of long-cells between silica-
	cell rows in costal zones
	Minority of long-cells in intercostal zones alternate with a
	silica- and short-cell pair; one to two rows of long-cells between
	silica-cell rows in costal zones
49.	Nodular-shaped silica-bodies occur in costal zones 50
	No nodular-shaped silica-bodies occur in costal zones 69
50.	Prickle-hairs occur
	Prickle-hairs absent 51
51.	Cells with papillae occur
	No cells with papillae occur
52.	Prickle-hairs absent
-0	More than one papilla per cell
53.	Short-cells and/or silica-cells occur in intercostal zones 54
	No short-cells and silica-cells occur in intercostal zones Trichoneura
54	Macro bairs present
54.	Macro-hairs present
55.	Majority of long-cells in intercostal zones alternate with a
55.	short- and silica-cell pair; one or two long-cell rows between
	silica-cell rows in costal zones Setaria flabellata
	Minority of long-cells in intercostal zones alternate with a
	short- and silica-cell pair; one row of long-cells between silica-
	cell rows in costal zones
56.	Long-cells in intercostal zones inflated with non-sinuous walls;
	majority of long-cells in intercostal zones alternate with a
	short- and silica-cell pair Setaria flabellata
	Long-cells in intercostal zones brick-shaped with sinuous walls;
	minority of long-cells in intercostal zones alternate with a
	short- and silica-cell pair
57.	The state of the s
	zones; no papillae
	No short- and silica-cell pairs in intercostal zones; papillae
50	sometimes present Andropogon schirensis var. angustifolius
58.	Interstomatal-cells taper sometimes a single row long-cells
	between silica-cell rows in costal zones . Monocymbium ceresiiforme
	Interstomatal-cells do not taper one or two long-cell rows

	between silica-cell rows in costal zones Setaria woodii
59.	Cells with papillae occur
	No cells with papillae 60
60.	Prickle-hairs in either costal or intercostal zones 61
	Prickle-hairs in costal as well as intercostal zones 62
61.	Prickle-hairs in costal zones Diplachne biflora Prickle-hairs in intercostal zones Brachiaria serrata
	Prickle-hairs in intercostal zones Brachiaria serrata
62.	Long-cells in intercostal zones brick-shaped with sinuous walls 63
04.	Long-cells in intercostal zones inflated with non-sinuous walls Setaria
	flabellata
63.	Macro-hairs occur
00.	Magna haire absent
64.	Prickle-hairs on margin of intercostal zones
01.	Prickle-hairs in centre of costal zones
65.	Interstomatal-cells longer than twice the length of the stomata
00.	Urochloa panicoides
	Interstomatal-cells shorter than twice the length of the stomata
	Trachypogon spicatus
66.	Basal cell of micro-hairs longer than distal cell Ctenium concinuum
00.	Basal cell of micro-hairs shorter than distal cell 67
67.	Interstomatal-cells longer than twice the length of stomata . 68
07.	Interstomatal-cells shorter than twice the length of stomata
	Trachypogon spicatus
68.	Interstomatal-cells horizontally tapered; sometimes a row of long-cells
00.	between silica-cell rows in costal zones . Monocymbium ceresiiforme
	Interstomatal-cells not horizontally tapered; one or two long-cell rows
	between silica-cell rows in costal zones Setaria woodii
69.	70
05.	Cells with papillae absent
70.	# 1
70.	More than one papilla per cell
71	
71.	Papillae on some interstomatal-cells only
70	Prickle-hairs occur
12.	No prickle-hairs occur
	No prickle-nairs occur Anatopogoti sentrensis var. angusty occur
73.	Basal cell of micro-hairs longer than stomata
2200	Basal cell of micro-hairs shorter than stomata. Trachypogon spicatus
74.	There-hans occur
	Titchic-mains absent
75.	
	No short-cells or silica-cells in intercostal zones
76.	Prickle-hairs in costal zones only
	Prickle-hairs in costal and intercostal zones
77.	Long-cells in intercostal zones brick-shaped with sinuous walls
	Diplachne biflora

Long-cells in intercostal zones inflated with non-sinuous walls Setaria pallide-fus	ca
	79
Long-cells in intercostal zones inflated with non-sinuous walls Setaria sphacela	
79. Basal cell of micro-hairs longer than distal cell Trichoneu grandiglum	ra
Basal cell of micro-hairs shorter than distal cell Andropogon eucom	
80. Prickle-hairs in costal zones only	31 33
81. Long-cells in intercostal zones brick-shaped with sinuous walls Long-cells in intercostal zones inflated with non-sinuous walls Setan pallide-fus	
82. Basal cell of micro-hairs longer than the stomata Aristida sp Basal cell of micro-hairs shorter than the stomata . Diplachne biflo	
83. Prickle-hairs on margins of costal zones	34 35
84. Macro-hairs in intercostal zones only Tristachya rehmann Macro-hairs, if present, on the margin of the leaf . Rhynchelytru repe	nii m
85. Interstomatal-cells horizontally tapered; sometimes a single row of long-cells between silica-cell rows in costal zones *Monocymbium ceresiiform*	
Interstomatal-cells not horizontally tapered; one or two long-	36
86. Macro-hairs, prickle-hairs and/or papillae occur; one, two or all three abovementioned appendages may in exceptional cases be absent	
Macro-hairs, prickle-hairs and papillae absent; in highly	
exceptional cases one, two or all three abovementioned	
appendages may be present Andropogon eucom	us
(Although these two grasses are easily distinguished from one another under a microscope, it is difficult to separate them in a key).	
87. Short-cells and/or silica-cells occur in intercostal zones 9	91 88
88. Long-cells in intercostal zones brick-shaped with sinuous walls	
Andropogon eucomus and Andropogon schirensis var. angustifolius or S Long-cells in intercostal zones inflated with non-sinuous walls Setaria pallide-fusca and Setaria sphacelata or S	

89.	
	centre of intercostal zones Setaria pallide-fusca
	Long-cells near costal zones do not differ significantly from
1.00.000	those in the centre of intercostal zones Setaria sphacelata
90.	Intercostal zones wider than five cell rows; papillae usually
	present
	present
91	Long-cell rows occur between silica-cell rows in costal zones . 92
51.	No long-cell rows occur between silica-cell rows in costal zones
	Monocymbium ceresiiforme
92.	Basal cell of micro-hairs longer than stomata 93
	Basal cell of micro-hairs shorter than stomata 94
93.	Silica-cells in costal zones wider than long-cells in intercostal
	zones
	Silica-cells in costal zones as wide as or narrower than long-
	cells in intercostal zones Loudetia simplex
94.	Majority of long-cells in intercostal zones alternate with short-
	and silica-cells individually or in pairs . Triraphis andropogonoides
	Minority of long-cells in intercostal zones alternate with short- or silica-cells individually or in pairs
05	Centre width of interstomatal-cells is larger than length of
95.	silica-cells in costal zones Setaria pallide-fusca
	Centre width of interstomatal-cells is smaller then length of
	silica-cells in costal zones Andropogon eucomus
96.	Silica-bodies in costal zones are irregular, cross-shaped, tall
	and narrow or square
	Silica-bodies in costal zones are saddle-shaped, round or oval 103
97.	Long-cell rows between silica-cell rows in costal zones present 98
	No long-cell rows between silica-cell rows in costal zones
	present
98.	Nodular-shaped silica-bodies in costal zones . Paspalum dilatatum
00	No nodular-shaped silica-bodies in costal zones Digitaria monodactyla Micro-hairs occur
99.	Micro-hairs occur
100	Prickle-hairs occur
100.	Prickle-hairs absent
101.	Cells with papillae occur
101.	Cells with papillae absent
102.	Subsidiary-cells of stomata dome-shaped Elyonurus argenteus
	Subsidiary-cells of stomata triangular Hemarthria altissima

103.	Long-cell rows between silica-cell rows in costal zones present No long-cell rows between silica-cell rows in costal zones
	present
104.	
	Micro-hairs absent Eragrostis gummiftua and Eragrostis plana or 105
105.	Majority of silica-bodies tall and narrow Eragrostis gummiflua
	Minority of silica-bodies tall and narrow Eragrostis plana
106.	Micro-hairs one-celled Sporobolus pyramidalis
	Micro-hairs two-celled
107.	Distal cell of micro-hairs longer than basal cell Eragrostis capensis
	Distal cell of micro-hairs shorter than basal cell Sporobolus pectinatus
108.	Micro-hairs occur
	Micro-hairs absent Eragrostis curvula
109.	Micro-hairs one-celled
	Micro-hairs two-celled
110.	Prickle-hairs occur
	Prickle-hairs absent
111.	Silica-cells in costal zones wider than length of stomata Microchloa
	caffra
	Silica-cells in costal zones as wide as or narrower then length
	of stomata
112.	Silica-cells occur in intercostal zones Cynodon dactylon
	No silica-cells in intercostal zones
113.	Prickle-hairs occur
	Prickle-hairs absent
114.	Prickle-hairs in costal and intercostal zones . Eragrostis heteromera
	Prickle-hairs in either costal or intercostal zones 115
115.	
	Prickle-hairs in costal zones
116.	Cells with papillae present
	No cells with papillae present
117.	Macro-hairs in intercostal zones
	Macro-hairs on margin of the leaf Tragus berteronianus Short-cells and/or silica-cells in intercostal zones
118.	Short cons unufor series
110	The black come of planet come and another another and another another and another another and another another another and another anot
119.	Digitaria monodactula
100	Macro-hairs on margin of the leaf Tragus berteronianus
120.	Macro-hairs on margin of the leaf Tragus berteronianus Basal cell of micro-hairs as long as or shorter than length of
120.	Macro-hairs on margin of the leaf Tragus berteronianus

121.	Short-cells and/or silica-cells present in intercostal zones 123
	No short-cells and silica-cells present in intercostal zones 122
122.	Macro-hairs in intercostal zones Digitaria monodactyla
	Macro-hairs on margin of the leaf Tragus berteronianus
123.	Basal cell of micro-hairs shorter than distal cell Eragrostis
	pseudo-sclerantha
	Basal cell of micro-hairs longer than distal cell 124
124.	Basal cell of micro-hairs shorter than stomata 125
	Basal cell of micro-hairs longer than stomata 126
125.	Three adjacent silica-cells occur in silica-cell rows of costal
	zones Eragrostis racemosa
	Three adjacent silica-cells in silica-cell rows of costal zones do
E38 11	not occur Pogonarthria squarrosa
126.	Interstomatal-cells shorter than stomata . Eragrostis chloromelas
	Interstomatal-cells longer than stomata Eragrostis heteromera

REFERENCES

- BAUMGARTNER, L. L. and A. C. MARTIN. 1939. Plant histology as an aid in squirrel food-habit studies. J. Wildl. Mgmt 3: 266-268.
- BORRILL, M. 1957. A morphologically distinct ecotype of *Dactylis glomerata* L. *Nature*, *ond*. 179: 544-545.
- CROKER, B. H. 1959. A method of estimating the botanical composition of the diet of sheep. N.Z. J. agric. Res. 2: 72-85.
- DAVIES, I. 1959. The use of epidermal characteristics for the identification of grasses in the leafy stage. J. Brit. Grassl. Soc. 14: 7-16.
- DUSI, J. L. 1949. Methods for the determination of food habits by plant microtechniques and histology and their application to cottontail rabbit food habits. J. Wildl. Mgmt 13: 295–298.
- DUVAL-JOUVE, M. J. 1875. Historie des Feuilles de Graminies. Ann. Sci. Nat. Bot. Ser. 6(1): 294-371.
- GOOSSENS, A. P. and J. J. THERON. 1934. An anatomical study of *Themeda triandra* Forsk. S. Afr. J. Sci. 31: 254-278.
- HERCUS, B. H. 1960. Plant cuticle as an aid to determine the diet of grazing animals. Proc. 8th Int. Grassl. Congr., (1960) Session IB. Grazing intake and behaviour studies: 443-447.
- LIVERSIDGE, R. 1970. Identification of grazed grasses using epidermal characters. *Proc. Grassld. Soc. S. Afr.* 5: 153-165.
- MARTIN, D. J. 1955. Features of plant cuticle. An aid to the analysis of the natural diet of grazing animals, with special reference to Scottish Hill Sheep. *Trans. and Proc. Bot. Soc. Edinb.* 36: 278–288.
- MARTIN, D. J. 1964. *In Crisp*, D. J.: Analysis of sheep diet utilizing plant epidermal fragments in faeces samples. Grazing in Terrestrial and Marine Environments. 173–188. Oxford: Blackwell Scient. Publ.
- McALLISTER, H. J. 1967. A quantative faecal analytical investigation of grazing preferences of impala (*Aepyceros m.melampus* Licht.). M.Sc. thesis. University of Natal, Pietermaritzburg.
- METCALFE, C. R. 1960. Anatomy of the Monocotyledones. I. Gramineae. Oxford: Clarendon Press.
- PRATT, H. 1932. L'Epiderme des Graminèes. Etude anatomique et systematique. Ann. Sci. Nat. Bot. Ser. 10(14): 117-324.
- STEWART, D. R. M. 1965. The epidermal characters of grasses with special reference to East African plains species. *Bot. Jahrb.* 84: 63–174.
- STEWART, D. R. M. 1967. Analysis of plant epidermis in faeces: a technique of studying the food preferences of grazing herbivores. J. appl. Ecol 4: 83-111.
- STEWART, D. R. M. 1971. Food preferences of an Impala herd. J. Wildl. Mgmt 35: 86-93.
- STEWART, D. R. M. and JOYCE STEWART. 1970. Food preference data by faecal analysis for African plains ungulates. Zool. afr. 5: 115-129.
- STORR, G. M. 1961. Microscopic analysis of faeces: a technique for ascertaining the diet of herbivorous mammals. *Aust. J. biol. Sci.* 14: 157–164.